

Bis(*N,N'*-diethyldiselenocarbamato)copper (II) and bis(*N,N'*-diethyldiselenocarbamato)zinc, reported by Barnard and Woodbridge,¹ were also prepared by the present method. The copper complex (black, m.p. 221°C) was recrystallized from chloroform and the zinc complex (yellow, m.p. 153–154°C) from benzene.

The yields of the complexes were almost quantitative. They were stored in a desiccator, over KOH and under nitrogen, in a cold room.

Bis(O-ethyldiselenocarbonato)nickel(II). Carbon diselenide (0.85 g) was added dropwise to a stirred and ice-cooled solution of potassium hydroxide (0.28 g) in ethanol, kept under nitrogen. This solution was added to a solution of NiCl₂·6H₂O (0.59 g) in water, with formation of a red precipitate. This was quickly extracted with chloroform, which on evaporation left black, glistening crystals of the complex.

Bis(N,N'-diethyldiselenocarbamato)nitrosylcobalt(II). Anhydrous cobalt(II) chloride (0.162 g) was dissolved in methanol, previously degassed with nitrogen, and the solution was saturated with nitrogen oxide. Sodium diethyl-diselenocarbamate (0.66 g) dissolved in methanol was added to the CoCl₂ solution under nitrogen and nitrogen oxide was again passed through the solution. A brown precipitate resulted which was filtered off, washed with cold methanol under nitrogen and recrystallized from chloroform.

Bis(N,N'-diethyldiselenocarbamato)nitrosyliron(II). FeCl₂·4H₂O (0.245 g) was dissolved in degassed methanol, and diethyl-diselenocarbamate (0.66 g) dissolved in methanol was added under nitrogen, with formation of a red precipitate. On saturation of the solution with nitrogen oxide, a brown precipitate separated. It was filtered off under nitrogen, washed with cold methanol, and crystallized from chloroform.

Acknowledgment. We thank Dr. Chr. Klíxbüll Jørgensen for a valuable discussion.

1. Barnard, D. and Woodbridge, D. T. *J. Chem. Soc.* 1961 2922.
2. Shankaranarayana, M. L. *Acta Chem. Scand.* To be published.
3. Jensen, K. A., Nielsen, P. H. and Engels-Henriksen, L. *Acta Chem. Scand.* To be published.
4. Jørgensen, C. K. *Mol. Phys.* 5 (1962) 485; Jørgensen, C. K. *Inorganic Complexes*, Academic, New York 1963, p. 166.

5. Lewis, J., Irving, R. J. and Wilkinson, G. *J. Inorg. Nucl. Chem.* 7 (1958) 32.

6. Jørgensen, C. K. *J. Inorg. Nucl. Chem.* 24 (1962) 1571.

Received November 24, 1967.

Mass Spectrometric and Gas Chromatographic Studies of *N*-Heptafluorobutyryl Derivatives of Peptide Methyl Esters

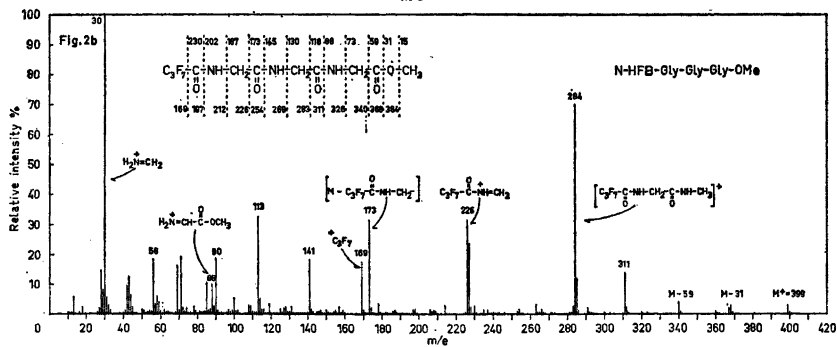
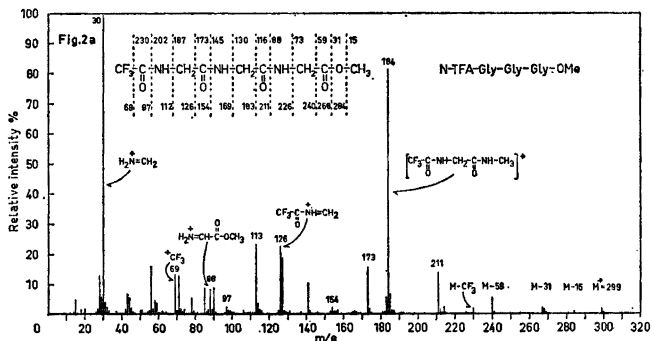
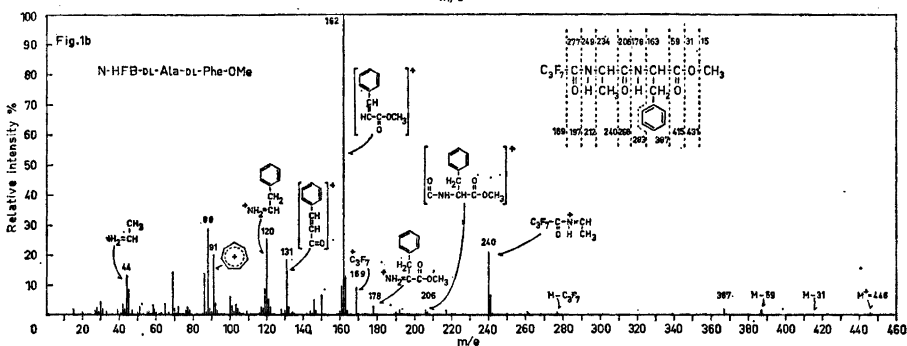
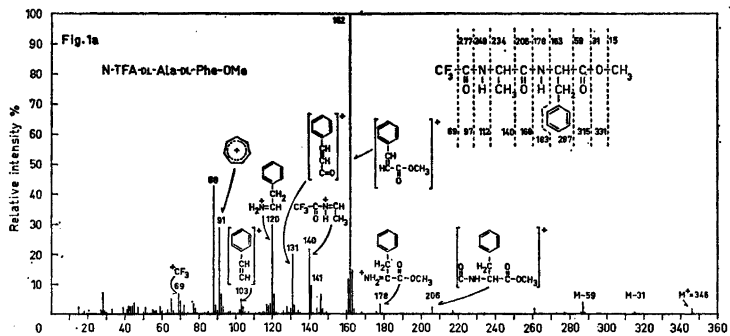
BENGT Å. ANDERSSON

Institute of Medical Biochemistry, University of Göteborg, Sweden

The *N*-trifluoroacetyl (*N*-TFA) derivatives of peptide methyl esters have been found useful in the direct mass spectrometric sequence analysis of peptides.¹⁻³

It has recently been found by Pollock⁴ that the *N*-heptafluorobutyryl (*N*-HFB) derivatives of butyl esters of amino acids have significantly shorter gas chromatographic retention times than the corresponding *N*-TFA derivatives. It was therefore of interest to study the mass spectrometric behaviour of the *N*-HFB derivatives of peptide methyl esters. Two peptide methyl esters, DL-Ala-DL-Phe-OMe and Gly-Gly-Gly-OMe, were acylated using the TFA and HFB anhydrides.⁵ The exchange of the trifluoroacetyl for the heptafluorobutyryl group reduced the gas chromatographic retention times on Carbowax 20M columns by approximately 50 %, and considerably lower temperatures were needed for the vapourization of the sample in the direct inlet system of the mass spectrometer. The mass spectra shown on Figs. 1 and 2 show the striking similarity in the fragmentation pattern between the TFA and HFB-derivatives, the only notable difference being that of 100 mass units in the *m/e* of ions containing the *N*-acyl group.

As peptide methyl esters *N*-acylated with long chain acyl groups give excellent mass spectra⁶ the mass spectrometric behaviour of peptide esters with higher *N*-perfluoroacyl groups is being studied.



This work was supported by *Statens Medicinska Forskningsråd*.

1. Andersson, C.-O. *Acta Chem. Scand.* **12** (1958) 1353.
2. Stenhagen, E. *Z. anal. Chem.* **181** (1961) 462.
3. Weygand, F., Prox, A., Fessel, H. H. and Sun, K. K. *Z. Naturforsch.* **20b** (1965) 1169.
4. Pollock, G. E. *Anal. Chem.* **39** (1967) 1194.
5. Weygand, F. and Geiger, R. *Chem. Ber.* **89** (1956) 647.
6. Barber, M., Jollès, E., Vilkas, E. and Lederer, E. *Biochem. Biophys. Res. Commun.* **18** (1965) 469.

Received November 18, 1967.

γ -Glutamyl-phenylalanine and γ -L-Glutamyl-L-tyrosine from Seeds of *Aubrietia deltoidea* DC.

P. OLESEN LARSEN and
HILMER SØRENSEN

Department of Organic Chemistry, The Royal Veterinary and Agricultural College, Copenhagen, Denmark

Previous communications from this laboratory reported results obtained during a systematic investigation of the free amino acids in species of Cruciferae.¹ In the course of these investigations the amino acid content in seeds of *Aubrietia deltoidea* DC. was determined by two-dimensional paper chromatography, and a spot was observed which could not be assigned to any amino acid previously identified in species of Cruciferae. The amino acid in question has now been isolated and identified as γ -L-glutamyl-L-tyrosine. In addition γ -glutamyl-phenylalanine has been isolated from the seeds. The latter compound was present in a concentration so small that it was not observed on the original paper chromatogram.

The fraction of acid amino acids from seeds of *A. deltoidea* DC. (1 kg, purchased from I. E. Ohlsen's Enke, Copenhagen) was

obtained by traditional methods including defatting with carbon tetrachloride, extraction with methanol:water, isolation of the total amino acid fraction on a strongly acid ion-exchange resin in the acid form with subsequent elution of the amino acids with ammonia, and isolation of the acid amino acids on a strongly basic ion-exchange resin in the acetate form with subsequent elution with acetic acid. Final purification was accomplished by ion-exchange chromatography on a strongly basic ion-exchange resin in the acetate form and by use of small ion-exchange columns and preparative paper chromatography as previously described.² Recrystallization from ethanol:water yielded γ -glutamyl-phenylalanine (4 mg, insufficient for the determination of optical rotation) and γ -L-glutamyl-L-tyrosine (101 mg, $[\alpha]_D^{25} + 26.8^\circ$ (c 1.1, H₂O). Lit. value $[\alpha]_D^{25} + 25.5^\circ$ (c 4, H₂O).³ The compounds were identified by comparison with authentic samples³ by use of infra-red absorption spectra and co-chromatography on paper. Furthermore, acid hydrolysis produced glutamic acid and phenylalanine, respectively tyrosine, as determined by co-chromatography on paper. The complete identity of the infra-red absorption spectrum of the isolated phenylalanine derivative with that of authentic γ -L-glutamyl-L-phenylalanine suggests L- (or more unlikely D-) configuration at both centers.

The presence of γ -glutamyl-tyrosine in seeds of *A. erubescens* Griseb. was established by paper chromatography. Traces of this compound may be present also in seeds of *Berteroa incana* (L.) DC. whereas the two γ -glutamyl derivatives have not been identified in any other species of Cruciferae investigated. A number of other γ -glutamyl derivatives are present in the crucifer *Lunaria annua* L.^{1,2}

γ -L-Glutamyl-L-phenylalanine and γ -L-glutamyl-L-tyrosine have been isolated previously from *Glycine max* (soybeans),³ *Lupinus angustifolius*, and *L. albus*.⁴ γ -L-Glutamyl-L-phenylalanine has been isolated also from *Allium cepa* where it occurs together with a number of other γ -glutamyl derivatives.⁵

The authors are indebted to Drs. C. J. Morris and J. F. Thompson for samples of γ -L-glutamyl-L-phenylalanine and γ -L-glutamyl-L-tyrosine.

1. Larsen, P. O. *Acta Chem. Scand.* **21** (1967) 1592.